

High-fat diet-induced obesity affects sulfurtransferases activity in mice hearts

DOMINIKA SZŁĘZAK¹, ANNA MISTERKA-KOZAKA², MONIKA MAJEWSKA-SZCZEPANIK³,
PATRYCJA BRONOWICKA-ADAMSKA¹

¹ Chair of Medical Biochemistry, Faculty of Medicine, Jagiellonian University Medical College,
Kraków, Poland

² Center for Biomedicine and Interdisciplinary Sciences, Faculty of Medicine,
Jagiellonian University Medical College, Kraków, Poland

³ Chair of Biomedical Sciences, Department of Medical Physiology, Faculty of Health Sciences,
Jagiellonian University Medical College, Kraków, Poland

Corresponding author: Dominika Szlęzak, Ph.D.
Chair of Medical Biochemistry, Faculty of Medicine
Jagiellonian University Medical College
ul. Kopernika 7, 31-034 Kraków, Poland
E-mail: dominika.szlęzak@uj.edu.pl

Abstract: Introduction: Obesity is a chronic disease with complex etiology, characterized by excessive accumulation of fat tissue, which poses a health hazard and increases, among others, risk of cardiovascular diseases. Hydrogen sulfide (H₂S), an endogenous gaseous transmitter, plays an important role in the cardiovascular system, affecting blood pressure and heart function.

Aim: The aim of this study was to determine the effect of high-fat diet-induced (HFD-induced) obesity to hydrogen sulfide metabolism in mice hearts.

Materials and Methods: The research material consisted of hearts collected from male wild-type C57BL/6 mice fed for 8 weeks with a normal diet (ND) or high-fat diet. *Ex vivo* analyses included testing the activity of selected sulfur enzymes (cystathionine gamma-lyase — CGL, 3-mercaptopyruvate sulfurtransferase — MPST, and thiosulfate sulfurtransferase — TST), determining the sulfane sulfur level, and examining the ability of tissues to produce hydrogen sulfide.

Results: Tissues collected from HFD-fed mice were characterized by a lack of CGL activity, increased TST activity, and higher levels of sulfane sulfur compared to group of ND mice. Additionally, the capacity to produce H₂S was higher in the hearts of HFD mice.

Conclusions: Our study shows that HFD-induced obesity affects hydrogen sulfide levels and sulfur enzyme activity in mice hearts. It can be speculated that the increased ability of HFD mice tissues to produce H₂S is a compensatory mechanism for obesity-induced changes. The conducted research indicates the key importance of sulfurtransferases, MPST and TST, in sulfur metabolism in the heart and indicates a new area for further analysis.

Keywords: hydrogen sulfide, thiosulfate sulfurtransferase, 3-mercaptopyruvate sulfurtransferase, high-fat diet, obesity, heart.

Submitted: 19-Dec-2025; **Accepted in the final form:** 30-Jan-2026; **Published:** 31-Mar-2026.



Introduction

Obesity is a disease characterized by the harmful effects of excess body fat on health. In clinical practice, the degree of obesity is usually assessed using the body mass index (BMI) — the ratio of body weight in kilograms to height in square meters (kg/m^2). [1] According to a report by the World Health Organization, in Europe alone, almost 60% of adults and nearly 8% of children are overweight or obese. [2] The accumulation of excess body fat leads to insulin resistance, glucose intolerance, type 2 diabetes, increased blood pressure, dyslipidemia, inflammation, and endothelial damage. All of these factors are considered cardiovascular risk mediators and may result in hypertension, coronary artery disease, heart failure, arrhythmia, atrial fibrillation, and premature sudden death. [3]

There are many causes of obesity, including unhealthy diet, lack of physical activity, genetic, hormonal, and psychological factors (such as stress or sleep disorders), socio-economic factors, and lack of education. [4] In the context of contemporary social changes, the issue of diet deserves special attention, including the increasingly widespread use of the so-called Western diet (WD). WD is high in calories and low in nutritional density. It includes nutrient-poor foods that are high in added sugar, salt, and saturated fat, such as fast food, sugary beverages, and highly processed food. [5] In animal research models, a high-fat diet (HFD) is considered a well-established model of the western diet. [6]

An inappropriate diet not only leads to obesity, but also to disturbances in the homeostasis of the organism and the development of oxidative/nitrosative stress, i.e. increased production of free radicals, such as reactive oxygen species and reactive nitrogen species. Physiological free radicals such as superoxide anion radical ($\bullet\text{O}_2^-$) and nitric oxide ($\bullet\text{NO}$), and their derivative — peroxynitrite (ONOO^-), are important modulators of many diseases, including cardiovascular disease. [7]

In the cardiovascular system, hydrogen sulfide (H_2S), an endogenously produced gaseous signaling molecule, has significant cardioprotective effects related to its vasodilatory properties. [8, 9] H_2S has been shown to protect against heart failure by stimulating angiogenesis, stimulating cardiac mitochondrial biogenesis, and activating endothelial nitric oxide synthase. [9] Therefore, hydrogen sulfide donors appear to be particularly useful therapeutic agents. The presence of sulfur atom/thiol group ($-\text{SH}$) in the structure of antihypertensive drugs from angiotensin-converting enzyme inhibitors (ACEI) group may intensify the cardioprotective effect of these medicine. [10]

In mammalian cells, H_2S is formed from L-cysteine in reactions catalyzed by three enzymes: cystathionine beta-synthase (CBS), cystathionine gamma-lyase (CGL), and 3-mercaptopyruvate sulfurtransferase (MPST). Additionally, hydrogen sulfide can be produced by the reduction of sulfane sulfur-containing compounds. [8] An important role in the metabolism of endogenous sulfur, is played by the enzyme thiosulfate sulfurtransferase (TST), which participates in the trans-sulfuration reaction. It catalyzes the transfer of sulfane sulfur from thiosulfate to thiol or cyanide, leading to the formation of persulfides or thiocyanates. [11] TST is a protein related evolutionarily to MPST and it is hypothesized that both may cooperate, including in eliminating the effects of oxidative stress. [12]

The aim of this study was to determine the effect of high-fat diet-induced obesity on the ability of mice heart tissue to produce hydrogen sulfide and on the activity of selected enzymes involved in sulfur metabolism. This work is a continuation of our previous experiments focusing on sulfur metabolism in tissues of animals with diet-induced obesity.

Materials and Methods

Animals and tissue collection

This examination was a continuation of our earlier study [13] evaluating the metabolism of sulfur-containing compounds in visceral adipose tissues collected from animals on normal (ND) or high-fat diet (approval no. 43/2017 of the 1st Local Ethics Committee for Animal Experiments in Cracow).

12-week-old wild-type C57BL/6 male mice from Jackson Laboratory (Bar Harbor, Maine, USA) were bred and kept under standardized conditions (light: dark cycle 12:12h, temperature 22°C, humidity 55%) with free access to food and water. The first group of animals ($n = 7$) were maintained on a standard/normal diet. To induce obesity, a second group of C57BL/6 mice ($n = 7$) were fed a high-fat diet D12492 from Research Diets, Inc. (New Brunswick, USA) *ad libitum* for 8 weeks. The energy sources in both diets and their caloric values are shown in Fig. 1. Mice body weight was monitored using a digital scale (Scout Pro Electronic Balance, Ohaus).

For tissue collection, the skin of mice anesthetized with a ketamine/xylazine cocktail (90 mg/kg body weight ketamine and 10 mg/kg body weight xylazine) was sterilized with 70% ethanol, and the thorax was incised to expose the heart. The entire heart was removed using sterile scissors and forceps. The tissue was placed in sterile tube, immediately frozen in liquid nitrogen, and stored at -80°C until further use.

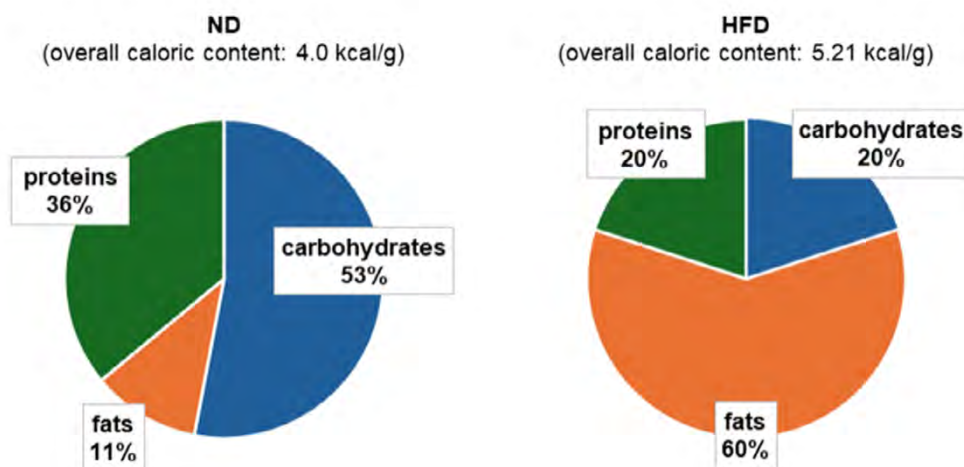


Fig. 1. Caloric composition of normal diet (ND) and high-fat diet (HFD) fed to animals.

Tissue homogenates

Sulfur enzyme activity, sulfane sulfur level, hydrogen sulfide production, and protein content were determined in mice heart homogenates prepared according to the procedures described in our previous work. [14] The obtained preparations were used immediately for further analyses.

Enzyme assay and determination of sulfane sulfur level

Cystathionine gamma-lyase activity was tested using the Matsuo and Greenberg method, [15] 3-mercaptopyruvate sulfurtransferase activity was determined according to the method of Valentine and Frankelfeld, [16] and thiosulfate sulfurtransferase activity was assayed by the Sörbo's method. [17] All these methods were modified by Wróbel *et al.* [18] The full composition of the reaction mixtures used and detailed procedures were described in our previous work. [19]

Sulfane sulfur was determined by Wood's cold cyanolysis method, [20] according to the procedure described previously. [19]

Protein content

Protein content in tissues was determined by the method of Lowry *et al.* [21] Crystalline bovine serum albumin was used as a standard.

H₂S production

The production of H₂S was examined by colorimetric method. [22, 23] The detailed composition of the reaction mixture and the experimental conditions were as described previously. [19]

Statistical analysis

All results are presented as arithmetic means $\pm$ standard error of the means. The Shapiro-Wilk test and Levene's test were used to assess the normal distribution of data and to analyze the homogeneity of variance. Statistical analysis was performed in StatSoft Statistica 13 software (Tibco, Palo Alto, California, USA) using the parametric Student's t test or the nonparametric Mann-Whitney test, as appropriate. Differences were considered statistically significant at the first level with a p value <0.05 , at the second level when $p < 0.01$, and at the third when $p < 0.001$.

Results

Animal parameters

Fig. 2 shows the mean body weights of the animals at the zero point of the experiment (12-week-old animals) and after 8 weeks of feeding with a normal diet or a high-fat diet (20-week-old animals). In the ND group, a 31% weight gain was recorded during the experiment. In contrast, the HFD group of mice showed a 75% increase in mean body weight, confirming the occurrence of obesity in these animals.

There were no statistically significant differences in the weight of hearts collected from animals fed the normal diet (mean weight 0.164 ± 0.003 g) and the high-fat diet (mean weight 0.158 ± 0.013 g) (data not shown).

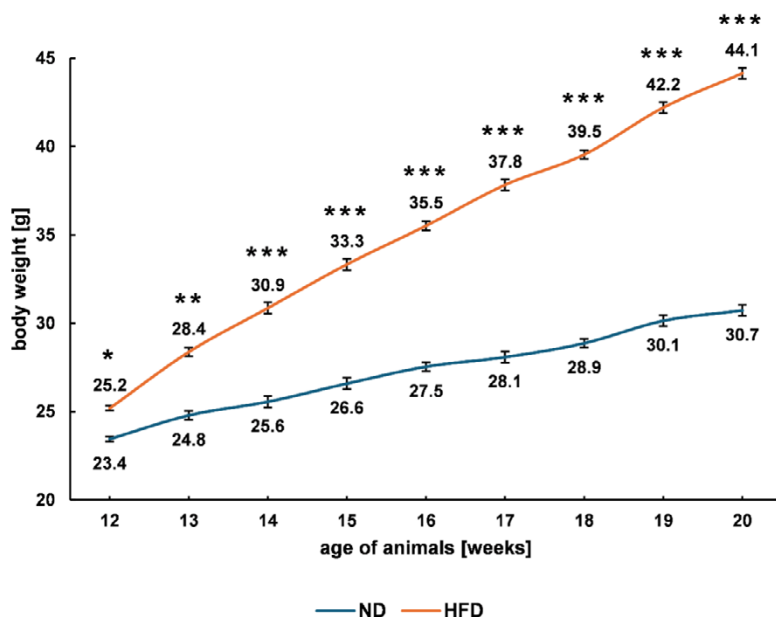


Fig. 2. Body weight changes of C57BL/6 mice fed a normal diet (ND) and a high-fat diet (HFD). Values are represented as arithmetic mean $\pm$ SEM of 7 animals. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ high fat diet (HFD) vs. normal diet (ND) (Mann-Whitney test).

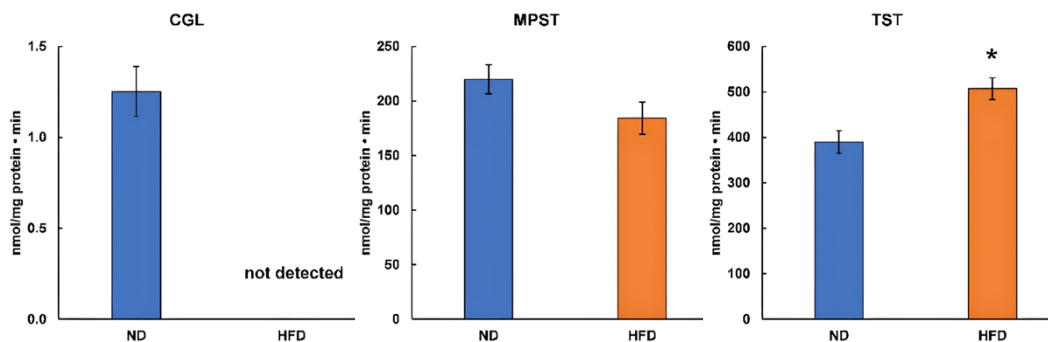


Fig. 3. The activity of cystathionine gamma-lyase (CGL), 3-mercaptopyruvate sulfurtransferase (MPST), and thiosulfate sulfurtransferase (TST) in mice hearts. Values are presented as the arithmetic mean $\pm$ SEM of 3 animals. Each value is the mean of 10–15 repeats. * $p < 0.05$ high fat diet (HFD) vs. normal diet (ND) (Mann-Whitney test).

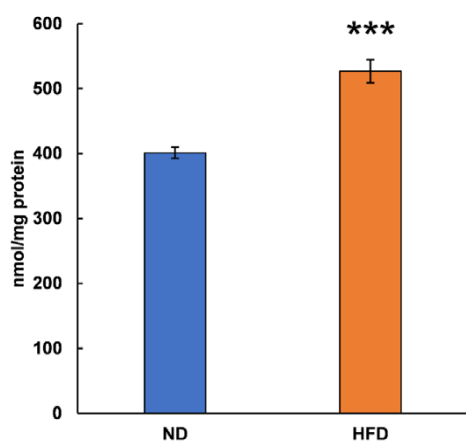


Fig. 4. The level of sulfane sulfur in mice hearts. Values are presented as the arithmetic mean $\pm$ SEM of 3 animals. Each value is the mean of 10–15 repeats. *** $p < 0.001$ high fat diet (HFD) vs. normal diet (ND) (Mann-Whitney test).

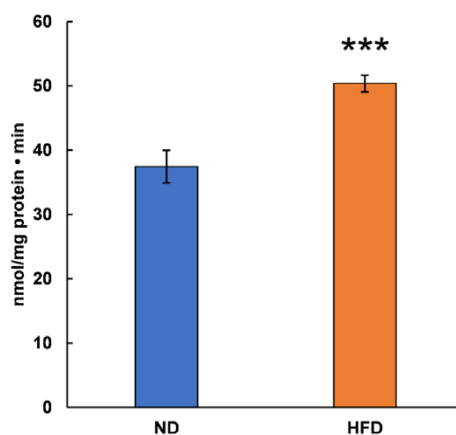


Fig. 5. Hydrogen sulfide (H₂S) production in mice hearts. Values are presented as the arithmetic mean $\pm$ SEM of 4 animals. Each value is the mean of 10–12 repeats. *** $p < 0.001$ high fat diet (HFD) vs. normal diet (ND) (Student's t test).

Enzymes activity and sulfane sulfur level

Enzymes activity in mice hearts are shown in Fig. 3. Low CGL activity was noticed in the tissues of animals fed a normal diet, while no CGL activity was detected in the HFD group. There were no statistically significant changes in MPST activity between the study groups. TST activity was significantly higher in the hearts of HFD mice compared to ND mice. Similarly, sulfane sulfur level increased significantly in the group of animals on the high-fat diet (Fig. 4).

H₂S production

The ability to generate hydrogen sulfide was confirmed in the tissues of animals from both study groups (Fig. 5). Hearts of HFD mice were characterized by significantly higher H₂S production.

Discussion

Obesity has been implicated as an indirect cause of heart disease for decades. Numerous studies in humans and animal models have shown that obesity leads to lipid accumulation in the heart, leading to cardiac steatosis. [24] Cardiac lipotoxicity, cardiomyocyte dysfunction and myocardial contractility dysfunction, is caused particularly by the deposition of saturated free fatty acids in heart cells. [25] The research by Alpert *et al.* [26] confirms the presence of left ventricular hypertrophy and impaired systolic and diastolic function in morbidly obese individuals, with the degree of impairment correlated with the duration of obesity. Left ventricular remodeling, along with increased heart rate and stroke volume, are characteristic of obesity and are thought to be

compensatory adaptations to increased adipose tissue mass. However, over time, they can lead to ischemic dilated cardiomyopathy. [27]

In this study, we focus on the changes in hydrogen sulfide metabolism induced in mice hearts by a high-fat diet. The results show that obesity determines the cardiac activity of sulfur enzymes. The new finding of this work is the important role of the TST enzyme in H₂S metabolism in the hearts of obese mice.

Cystathionine gamma-lyase has been the subject of interest of many research groups for many years as the main enzyme responsible for hydrogen sulfide production in the circulatory system. [28, 29, 30] CGL catalyzes the formation of H₂S from L-cysteine with the release of pyruvate. It can also induce the formation of thiosulfate from L-cystine and is responsible for the catabolism of cystathionine occurring on the pathway of L-cysteine formation from L-methionine. [31] Analysis of enzyme activity showed that the low CGL activity in the hearts of mice fed normal diet was reduced to undetectable level in HFD-fed animals. These results are consistent with immunohistochemical evaluation of CGL performed by Peh *et al.*, [32] which showed a significant reduction or absence of CGL in aortic endothelial cells in C57BL/6 mice fed HFD for 16 weeks relative to mice on a standard diet. However, this change was not associated with a reduced ability of these tissues to produce hydrogen sulfide. These results indicate that other enzymes besides CGL are involved in maintaining the H₂S pool in heart cells.

Studies by various groups of scientists indicate the significant importance of 3-mercaptopyruvate sulfurtransferase in maintaining the proper functioning of the cardiovascular system. It has been shown that knockout of the MPST enzyme causes hypertension and cardiac hypertrophy in 18-month-old mice compared to younger animals. [33] Another study [34] showed that coronary rat homogenates produce larger amounts of hydrogen sulfide through the enzymatic pathway using MPST (desulfuration of 3-mercaptopyruvate leading to the formation of sulfane sulfur-rich compounds [35]) than through CGL. Our previous studies [14] in the rat model of hypertension also confirm that 3-mercaptopyruvate sulfurtransferase plays a major role in H₂S production in the heart. However, studies conducted on mice with HFD-induced metabolic syndrome did not show any changes in MPST expression at the protein level in the hearts of these animals compared to control mice. [36] Similarly, no changes were observed in the cardiac levels of free hydrogen sulfide and sulfane sulfur. The same study [36] and study by Katsouda *et al.* [37] showed that MPST knockout in mice resulted in increased pathological changes associated with metabolic syndrome (increased body weight, impaired glucose tolerance) and contributed to vascular inflammation. On the other hand, reports on MPST expression in other tissues of the body are contradictory — Li *et al.* [38] report that MPST expression in the liver at protein level increases in C57BL/6 mice fed a high-fat diet for 8 weeks, while Peh *et al.* [32] indicate a decrease in the hepatic MPST protein level in the same animal model after 8, 12, and 16 weeks of HFD application.

In this study, we did not observe significant changes in MPST activity in mice hearts under the influence of the high-fat diet. However, the obtained results suggest a tendency towards reduced activity of this enzyme in HFD-fed mice, which may be related to its antioxidant potential. Cysteine (Cys) located in the catalytic center of the enzyme and having reducing properties, plays a particularly important antioxidant role. The presence of Cys allows the reduction of oxidizing compounds such as reactive oxygen species, but also causes the inactivation of MPST. [39]

Another important enzyme involved in sulfur metabolism is mitochondrial thiosulfate sulfurtransferase. There are reports suggesting that persistent oxidative stress (especially in the

mitochondrial region) may lead to the induction of TST, which (in cooperation with MPST) regulates the level of sulfane sulfur, reduced glutathione or thioredoxin, causing an antioxidant effect. [12] In the current experiment, we observed a significant increase in cardiac TST activity under the influence of HFD, which may result from the activation of the mentioned antioxidant mechanism. Similarly, the level of sulfane sulfur was significantly higher in the HFD mice group. It is worth emphasizing that TST, although not classified as an H₂S-producing enzyme, catalyzes the reaction of thiosulfate with glutathione to produce hydrogen sulfide. [31] The presented results confirm that TST activity plays an important role in the metabolism of hydrogen sulfide in obese mice.

The increased bioavailability of sulfane sulfur (direct hydrogen sulfide precursor [40]) in HFD mice tissues explains the high capacity of these tissues to generate H₂S and may be due to its physicochemical properties. Elemental sulfur is characterized by high solubility in lipids. [41] Studies on obesity, both in humans and using animal models, [42, 43, 44] confirm that the heart is an important absorber of fatty acids from the plasma due to its high energy demand. At the same time, uptake of fatty acids from the circulation prevents their harmful effects on other tissues such as the pancreas. [43]

A limitation of our study is that we used only a male research model. Since there are reports of gender differences in cardiovascular risk, [45] further studies should be performed with a female research model. In addition, studies using an animal model have some inherent limitations, including the number of animals in each experimental group. The relatively small number of *n* in our research groups can be considered a limitation of this study.

Conclusions

In summary, high-fat diet-induced obesity affects hydrogen sulfide levels and sulfur enzyme activity in mice hearts. The increased capacity of heart tissue from mice fed a high-fat diet to produce H₂S could be interpreted as a compensatory mechanism aimed at counteracting obesity-induced changes, such as metabolic disorders or oxidative stress. Enzymatic analysis highlights the critical role of sulfurtransferases, MPST and TST, in cardiac sulfur metabolism, suggesting their key role in regulating hydrogen sulfide levels and maintaining redox balance in cardiac tissue. This demonstrates a potential role for these enzymes in cardiac adaptation to obesity and provides a new area for further investigation to understand the mechanisms behind obesity-related cardiovascular disease.

Declaration of competing interest

The authors declare no competing interests related to the present study.

Authorship contribution statement

D.S.: conceptualization, data curation, formal analysis, investigation, methodology, visualization, writing — original draft, writing — review and editing; A.M.-K.: data curation, formal analysis; M.M.-S.: data curation, funding acquisition, methodology; P.B.-A.: conceptualization, data curation, funding acquisition, project administration, supervision, writing — review and editing.

Funding

This work was supported by the Polish Ministry of Science and Higher Education (grants no. N41/DBS/000938 and K/ZDS/007123).

References

1. González-Muniesa P, Martínez-González M.A., Hu F.B., et al.: Obesity. *Nat Rev Dis Primers*. 2017; 3: 17034. <https://doi.org/10.1038/nrdp.2017.34>.
2. World Health Organization. European Regional Obesity Report 2022. Copenhagen, Denmark, 2022.
3. Piché M.E., Tchernof A., Després J.P.: Obesity Phenotypes, Diabetes, and Cardiovascular Diseases. *Circ Res*. 2020; 126 (11): 1477–1500. <https://doi.org/10.1161/CIRCRESAHA.120.316101>.
4. Chatterjee A., Gerdes M.W., Martinez S.G.: Identification of Risk Factors Associated with Obesity and Overweight — A Machine Learning Overview. *Sensors (Basel)*. 2020; 20 (9): 2734. <https://doi.org/10.3390/s20092734>.
5. Clemente-Suárez V.J., Beltrán-Velasco A.I., Redondo-Flórez L., Martín-Rodríguez A., Tornero-Aguilera J.F.: Global Impacts of Western Diet and Its Effects on Metabolism and Health: A Narrative Review. *Nutrients*. 2023; 15 (12): 2749. <https://doi.org/10.3390/nu15122749>.
6. Pakiet A., Jakubiak A., Mierzejewska P., et al.: The Effect of a High-Fat Diet on the Fatty Acid Composition in the Hearts of Mice. *Nutrients*. 2020; 12 (3): 824. <https://doi.org/10.3390/nu12030824>.
7. Griendling K.K., Touyz R.M., Zweier J.L., et al.: Measurement of Reactive Oxygen Species, Reactive Nitrogen Species, and Redox-Dependent Signaling in the Cardiovascular System: A Scientific Statement From the American Heart Association. *Circ Res*. 2016; 119 (5): e39–e75. <https://doi.org/10.1161/RES.000000000000110>.
8. Kimura H.: Hydrogen sulfide and polysulfides as signaling molecules. *Proc Jpn Acad Ser B Phys Biol Sci*. 2015; 91 (4): 131–159. <https://doi.org/10.2183/pjab.91.131>.
9. Wu D., Hu Q., Zhu D.: An Update on Hydrogen Sulfide and Nitric Oxide Interactions in the Cardiovascular System. *Oxid Med Cell Longev*. 2018; 2018: 4579140. <https://doi.org/10.1155/2018/4579140>.
10. Terzuoli E., Monti M., Vellecco V., et al.: Characterization of zofenoprilat as an inducer of functional angiogenesis through increased H₂S availability. *Br J Pharmacol*. 2015; 172 (12): 2961–2973. <https://doi.org/10.1111/bph.13101>.
11. Casin K.M., Calvert J.W.: Harnessing the Benefits of Endogenous Hydrogen Sulfide to Reduce Cardiovascular Disease. *Antioxidants (Basel)*. 2021; 10 (3): 383. <https://doi.org/10.3390/antiox10030383>.
12. Nakajima T.: Roles of Sulfur Metabolism and Rhodanese in Detoxification and Anti-Oxidative Stress Functions in the Liver: Responses to Radiation Exposure. *Med Sci Monit*. 2015; 21: 1721–1725. <https://doi.org/10.12659/MSM.893234>.
13. Bronowicka-Adamska P., Szlęzak D., Bentke-Imiolek A., Kaszuba K., Majewska-Szczepanik M.: The modulation of low molecular weight sulfur compounds levels in visceral adipose tissue of TLR2-deficient mice on a high-fat diet. *Biochimie*. 2025; 232: 66–73. <https://doi.org/10.1016/j.biochi.2025.01.008>.
14. Szlęzak D., Hutsch T., Ufnal M., Wróbel M.: Heart and kidney H₂S production is reduced in hypertensive and older rats. *Biochimie*. 2022; 199: 130–138. <https://doi.org/10.1016/j.biochi.2022.04.013>.
15. Matsuo Y., Greenberg D.M.: A crystalline enzyme that cleaves homoserine and cystathionine. I. Isolation procedure and some physicochemical properties. *J Biol Chem*. 1958; 230 (2): 545–560.
16. Valentine W.N., Frankenfeld J.K.: 3-Mercaptopyruvate sulfurtransferase (EC 2.8.1.2): a simple assay adapted to human blood cells. *Clin Chim Acta*. 1974; 51 (2): 205–210. [https://doi.org/10.1016/0009-8981\(74\)90031-x](https://doi.org/10.1016/0009-8981(74)90031-x).
17. Sörbo B.: Rhodanese. [In] *Methods in Enzymology*. S.P. Colowick, N.O. Kaplan (eds.). Academic Press, New York, 1955; 334–337.

18. Wróbel M., Jurkowska H., Sliwa L., Srebro Z.: Sulfurtransferases and cyanide detoxification in mouse liver, kidney, and brain. *Toxicol Mech Methods*. 2004; 14 (6): 331–337. <https://doi.org/10.1080/15376520490434683>.
19. Szlęzak D., Bronowicka-Adamska P., Hutsch T., Ufnal M., Wróbel M.: Hypertension and aging affect liver sulfur metabolism in rats. *Cells*. 2021; 10 (5): 1238. <https://doi.org/10.3390/cells10051238>.
20. Wood J.L.: Sulfanesulfur. *Methods Enzymol*. 1987; 143: 25–29. [https://doi.org/10.1016/0076-6879\(87\)43009-7](https://doi.org/10.1016/0076-6879(87)43009-7).
21. Lowry O.H., Rosebrough N.J., Farr A.L., Randall R.J.: Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951; 193 (1): 265–275.
22. Stipanuk M.H., Beck P.W.: Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. *Biochem J*. 1982; 206 (2): 267–277. <https://doi.org/10.1042/bj2060267>.
23. Collin M., Anuar F.B.M., Murch O., Bhatia M., Moore P.K., Thiernemann C.: Inhibition of endogenous hydrogen sulfide formation reduces the organ injury caused by endotoxemia. *Br J Pharmacol*. 2005; 146 (4): 498–505. <https://doi.org/10.1038/sj.bjp.0706367>.
24. Ghosh S., Sulistonyingrum D.C., Glier M.B., Verchere C.B., Devlin A.M.: Altered glutathione homeostasis in heart augments cardiac lipotoxicity associated with diet-induced obesity in mice. *J Biol Chem*. 2011; 286 (49): 42483–42493. <https://doi.org/10.1074/jbc.M111.304592>.
25. Yarmohammadi F., Hayes A.W., Karimi G.: Natural and chemical compounds as protective agents against cardiac lipotoxicity. *Biomed Pharmacother*. 2022; 145: 112413. <https://doi.org/10.1016/j.biopha.2021.112413>.
26. Alpert M.A., Lambert C.R., Panayiotou H., et al.: Relation of duration of morbid obesity to left ventricular mass, systolic function, and diastolic filling, and effect of weight loss. *Am J Cardiol*. 1995; 76 (16): 1194–1197. [https://doi.org/10.1016/s0002-9149\(99\)80338-5](https://doi.org/10.1016/s0002-9149(99)80338-5).
27. Szczepaniak L.S., Victor R.G., Orci L., Unger R.H.: Forgotten but not gone: the rediscovery of fatty heart, the most common unrecognized disease in America. *Circ Res*. 2007; 101 (8): 759–767. <https://doi.org/10.1161/CIRCRESAHA.107.160457>.
28. Yan H., Du J., Tang C.: The possible role of hydrogen sulfide on the pathogenesis of spontaneous hypertension in rats. *Biochem Biophys Res Commun*. 2004; 313 (1): 22–27. <https://doi.org/10.1016/j.bbrc.2003.11.081>.
29. Yang G., Wu L., Jiang B., et al.: H₂S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. *Science*. 2008; 322 (5901): 587–590. <https://doi.org/10.1126/science.1162667>.
30. Wijerathne C.U.B., Madduma Hewage S., Siow Y.L., O K.: Kidney ischemia reperfusion decreases hydrogen sulfide and increases oxidative stress in the heart. *Biomolecules*. 2020; 10 (11): 1565. <https://doi.org/10.3390/biom10111565>.
31. Wang R.: Physiological implications of hydrogen sulfide: a whiff exploration that blossomed. *Physiol Rev*. 2012; 92 (2): 791–896. <https://doi.org/10.1152/physrev.00017.2011>.
32. Peh M.T., Anwar A.B., Ng D.S., Atan M.S., Kumar S.D., Moore P.K.: Effect of feeding a high fat diet on hydrogen sulfide (H₂S) metabolism in the mouse. *Nitric Oxide*. 2014; 41: 138–145. <https://doi.org/10.1016/j.niox.2014.03.002>.
33. Peleli M., Bibli S.I., Li Z., et al.: Cardiovascular phenotype of mice lacking 3-mercaptopyruvate sulfurtransferase. *Biochem Pharmacol*. 2020; 176: 113833. <https://doi.org/10.1016/j.bcp.2020.113833>.
34. Kuo M.M., Kim D.H., Jandu S., et al.: MPST but not CSE is the primary regulator of hydrogen sulfide production and function in the coronary artery. *Am J Physiol Heart Circ Physiol*. 2016; 310: H71–H79. <https://doi.org/10.1152/ajpheart.00574.2014>.
35. Pedre B., Dick T.P.: 3-Mercaptopyruvate sulfurtransferase: an enzyme at the crossroads of sulfane sulfur trafficking. *Biol Chem*. 2020; 402 (3): 223–237. <https://doi.org/10.1515/hsz-2020-0249>.
36. Zampas P., Li Z., Katsouda A., et al.: Protective role of 3-mercaptopyruvate sulfurtransferase (MPST) in the development of metabolic syndrome and vascular inflammation. *Pharmacol Res*. 2025; 211: 107542. <https://doi.org/10.1016/j.phrs.2024.107542>.
37. Katsouda A., Valakos D., Dionellis V.S., et al.: MPST sulfurtransferase maintains mitochondrial protein import and cellular bioenergetics to attenuate obesity. *J Exp Med*. 2022; 219 (7): e20211894. <https://doi.org/10.1084/jem.20211894>.

38. Li M., Xu C., Shi J., et al.: Fatty acids promote fatty liver disease via the dysregulation of 3-mercaptopyruvate sulfurtransferase/hydrogen sulfide pathway. *Gut*. 2018; 67 (12): 2169–2180. <https://doi.org/10.1136/gutjnl-2017-313778>.
39. Nagahara N.: Multiple role of 3-mercaptopyruvate sulfurtransferase: antioxidative function, H₂S and polysulfide production and possible SO_x production. *Br J Pharmacol*. 2018; 175 (4): 577–589. <https://doi.org/10.1111/bph.14100>.
40. Bilska-Wilkosz A., Głowacka K., Kocemba-Pilarczyk K., Marcykiewicz B., Górny M., Iciek M.: α -Lipoic acid increases the viability of nephrocytes and elevates sulfane sulfur level in plasma of patients with chronic kidney disease. *Folia Med Cracov*. 2024; 64 (1): 39–52. <https://doi.org/10.24425/fmc.2024.150140>.
41. Toohey J.I.: Possible involvement of sulfane sulfur in homocysteine-induced atherosclerosis. *Med Hypotheses*. 2001; 56 (2): 259–261. <https://doi.org/10.1054/mehy.2000.1209>.
42. Aasum E., Hafstad A.D., Severson D.L., Larsen T.S.: Age-dependent changes in metabolism, contractile function, and ischemic sensitivity in hearts from db/db mice. *Diabetes*. 2003; 52 (2): 434–441. <https://doi.org/10.2337/diabetes.52.2.434>.
43. Peterson L.R., Herrero P., Schechtman K.B., et al.: Effect of obesity and insulin resistance on myocardial substrate metabolism and efficiency in young women. *Circulation*. 2004; 109 (18): 2191–2196. <https://doi.org/10.1161/01.CIR.0000127959.28627.F8>.
44. Marín-Royo G., Ortega-Hernández A., Martínez-Martínez E., et al.: The Impact of Cardiac Lipotoxicity on Cardiac Function and Mirnas Signature in Obese and Non-Obese Rats with Myocardial Infarction. *Sci Rep*. 2019; 9 (1): 444. <https://doi.org/10.1038/s41598-018-36914-y>.
45. Baranowska A., Skowron B., Ciesielczyk K., Domagała J., Thor P.J.: Experimental gender related obesity effect of diet. *Folia Med Cracov*. 2016; 56 (1): 49–60.